



Formulation and evaluation of hydrogel beads of flecainide

Penaka Nomika Reddy¹, P Venkata Anudeep*¹, Venugopalaiah Penabaka¹, Yadala Prapurna Chandra²

¹Department of Pharmaceutics, Ratnam Institute of Pharmacy, Pidathapolur (V & P), Muthukur (M), SPSR Nellore District-524 346.

²Department of Pharmacology, Ratnam Institute of Pharmacy, Pidathapolur (V & P), Muthukur (M), SPSR Nellore District-524 346.

Article History:

Received on: 23 Jul 2024
Revised on: 27 Sep 2024
Accepted on: 30 Sep 2024

Keywords:

Nanoparticles,
Anti-Cancer,
Paclitaxel,
Melanoma.

Abstract



This study focuses on creating and assessing hydrogel beads containing Flecainide. Hydrogels are polymer networks that absorb and retain significant amounts of water. Within this network, hydrophilic groups become hydrated in aqueous environments, forming a hydrogel structure. The primary goal was to evaluate the formulation of hydrogel beads with Flecainide. Preliminary studies, including solubility and UV analysis, confirmed the formulation's requirements. FTIR spectra indicated no interaction between Flecainide and the polymers, suggesting that the distribution of Flecainide within the beads was appropriate and within acceptable limits. Additionally, the study demonstrated that as the polymer concentration increases, the amount of medication released decreases. The Flecainide hydrogel beads exhibited controlled and extended drug release in vitro. The dissolution data for the optimal formulation (F12) were analyzed using three kinetic models: the Higuchi and Korsmeyer-Peppas equations, zero-order, and first-order kinetics. The r^2 value for the optimized formulation F12 is 0.974, indicating compliance with zero-order release kinetics. Furthermore, the Korsmeyer-Peppas analysis supports the mechanism of drug release. For formulation F12, the "n" value is 1.021, indicating a supercase transport mechanism.

*Corresponding Author

Name: P Venkata Anudeep
Phone: +91 81436 59012
Email: anudeepadavala9@gmail.com

eISSN: 2583-5254

DOI: <https://doi.org/10.26452/ijebr.v3i4.669>



Production and hosted by
Pharmasprings.com
© 2024 | All rights reserved

INTRODUCTION

Hydrogels are polymeric networks that can absorb and hold vast amounts of water. A hydrogel structure is created by the hydrophilic groups in the polymeric network hydrating in aqueous surroundings. It can alternatively be described as a polymeric material that has the ability to swell and retain a significant amount of water inside its structure despite not dissolving in water. They resemble natural tissue because they are somewhat flexible due to their high water content. Hydrogels are resistant to dissolution because of

crosslinks between network chains, and they can absorb water because of hydrophilic functional groups attached to the polymeric backbone [1]. A hydrogel comprises a three-dimensional (3D) network of hydrophilic polymers that have been crosslinked chemically or physically to allow the polymer chains to swell in water and retain a significant amount of water while retaining their structure. In the beginning, Wichterle and Lím reported on hydrogels. To qualify as a hydrogel, a material's weight (or volume) must include water at least 10% of the total. Hydrogels possess an extraordinarily close-to-natural tissue degree of elasticity, primarily attributed to their high water content. The hydrophilicity of the network is explained by the presence of hydrophilic groups such as -NH₂, -COOH, -OH, -CONH₂, -CONH-, and -SO₃H. Hydrogels undergo a significant volume phase change, also known as a gel-sol phase transition, in response to particular physical and chemical stimuli [2].

METHODOLOGY

PREFORMULATION STUDY:

the potassium bromide pellet method, the FTIR spectra of Flecainide and formulation comprising all polymers were determined in the wavelength range of 4,000 to 400 cm⁻¹. The process involved distributing a sample of potassium bromide and compressing it into discs using a hydraulic press set to five tonnes of pressure for five minutes. The spectrum was acquired after inserting the pellet into the light path [4].

Methodology

Formulation of Hydrogel Beads:

Ionotropic gelation is the process used to make hydrogel beads. A precise amount of polymer was dissolved in 25 milliliters of purified water and mixed to create a dispersion. The drug was added to the dispersion mentioned above and mixed once more to ensure uniform distribution and create a homogenous mixture. Using a 23G syringe needle, the mixture was extruded into a 1% w/v calcium chloride solution. The beads were left in the same solution for thirty minutes to increase their mechanical strength. After the beads were created, they were separated, given a water wash,

Table 1 Formulation Design for Flecainide hydrogel beads

Ingredients	F1	F2	F3	F4	F5	F6	F7	F8	F9	F10	F11	F12
Flecainide	150	150	150	150	150	150	150	150	150	150	150	150
Sodium Alginate	150	300	450	600	150	300	450	600	150	300	450	600
Sodium CMC	150	300	450	600	-	-	-	-	-	-	-	-
HPMC K4M	-	-	-	-	150	300	450	600	-	-	-	-
Carbopol	-	-	-	-	-	-	-	-	150	300	450	600
Calcium chloride (%)	2	2	2	2	2	2	2	2	2	2	2	2

FTIR study:

Fourier transform infrared spectroscopy was used in drug-polymer compatibility studies to verify that the drug's entrapment within polymeric systems is solely a physical process and that there is no ongoing interaction between the drug and polymer combination. To verify the identity of the drug and identify any interactions between it and the excipients, FTIR absorption spectra of the pure drug, all of the polymers utilized, and the combination of drug and polymers were obtained [3]. The drug's compatibility with the formulation was verified using FTIR spectrum analysis. Using the Shimadzu FT-IR 8300 spectrophotometer and

and left to dry overnight at room temperature [5].

Evaluation of Hydrogel Beads

Surface Morphology (SEM)

Particle size distribution, surface topography, texture, and the morphology of shattered or sectioned surfaces have all been studied using scanning electron microscopy. SEM is arguably the most widely used technique for characterizing drug delivery systems since it is easy to use and requires little material preparation [6]. SEM research was conducted on a JEOL JSM T-330A scanning microscope. An electron microscope brass stub was coated with dry flecainide gel

beads using an ion sputter. Random stub scanning was used to capture images of the flecainide hydrogel beads [7].

Percentage Yield

The % practical yield of flecainide hydrogel beads was computed to determine the efficiency or yield percentage of any given process, which aids in selecting the most suitable manufacturing technique [8]. The weight of Flecainide beads recovered from each batch divided by the total starting material was used to compute the practical yield. Using the formula, the % yield of prepared flecainide beads was ascertained.

Percentage yield

$$= \frac{\text{Practical yield}}{\text{Theoretical yield}} \times 100$$

Drug Content

A porcelain mortar and pestle were used to smash 40 mg of beads, which were then dispersed in an appropriate solvent to measure the beads' drug concentration and encapsulation efficiency. It was filtered after the dispersion was allowed overnight for 24 hours and sonicated for 15 minutes. A UV-visible spectrophotometer with a λ_{max} of 232 nm was used to measure the drug content of a 1 ml sample diluted with an appropriate solvent. Each formulation's medication content was stated as mg / 200 mg of gel beads [9].

Drug Entrapment Efficiency

The following formula was used to calculate the drug entrapment efficiency of the produced beads [10].

$$EE (\%) = \frac{\text{Actual Drug Content}}{\text{Theoretical Drug Content}} \times 100$$

In-vitro dissolution studies

Procedure for In-vitro Dissolution Study

The paddle method of USP XXIII equipment II was utilized to ascertain the release rate of Flecainide Hydrogel beads. The dissolution test was run with 900 milliliters of 0.1N HCL for two hours and a 6.8 pH buffer for ten hours at 50 rpm, $37 \pm 0.5^\circ\text{C}$. For the investigation, 40 mg of flecainide hydrogel beads were employed [11]. Five milliliters of the sample solution were taken out of the dissolving device for up to twelve hours at different times

(hourly). A new dissolution medium was added to the samples. After filtering the samples, the absorbance at $\lambda_{\text{max}} 232\text{nm}$ was calculated. A cumulative percentage drug release versus time plot was used to analyze the dissolution profiles of the formulations. Kinetic treatment was also applied to the collected data [12] to comprehend the release process.

Mathematical modeling for drug release profile

Zero-order kinetics: It explains the mechanism wherein the drug release rate is unaffected by concentration.

$$Qt = Q_0 + K_0t$$

First order kinetics

The drug release from systems where the release rate depends on concentration is described [13].

$$\text{Log } Qt = \frac{\text{Log } Q_0 K_1 t}{2.303}$$

Higuchi model

It explains how the square root of time determines the proportion of drug release from a matrix.

$$Mt/M_\infty = K_H t^{1/2}$$

Korsmeyer-Peppas model (Power law)

The potent law effectively characterizes the release of drugs from slabs, cylinders, and spheres by stating that the fractional amount of drug release is exponentially related to the release time [14]

$$\frac{Mt}{M_\infty} = Ktn$$

$$\text{Log}\left(\frac{Mt}{M_\infty}\right) = \log K + n \log t$$

Stability Conditions

For one month and three months, the following temperatures were used to examine the stability of tablets containing Flecainide.

1. Extended testing period: 1 month at 25°C and 60% relative humidity (3 months)
2. Quick testing: 1 month at 40°C and 75% relative

humidity (3months) Estimated parameters: drug content [15].

Drug polymer interaction study

It was discovered from the spectra of Flecainide, Flecainide and blank beads, and Flecainide and polymer mixture that all of Flecainide's distinctive

RESULTS AND DISCUSSION

PREFORMULATION STUDY

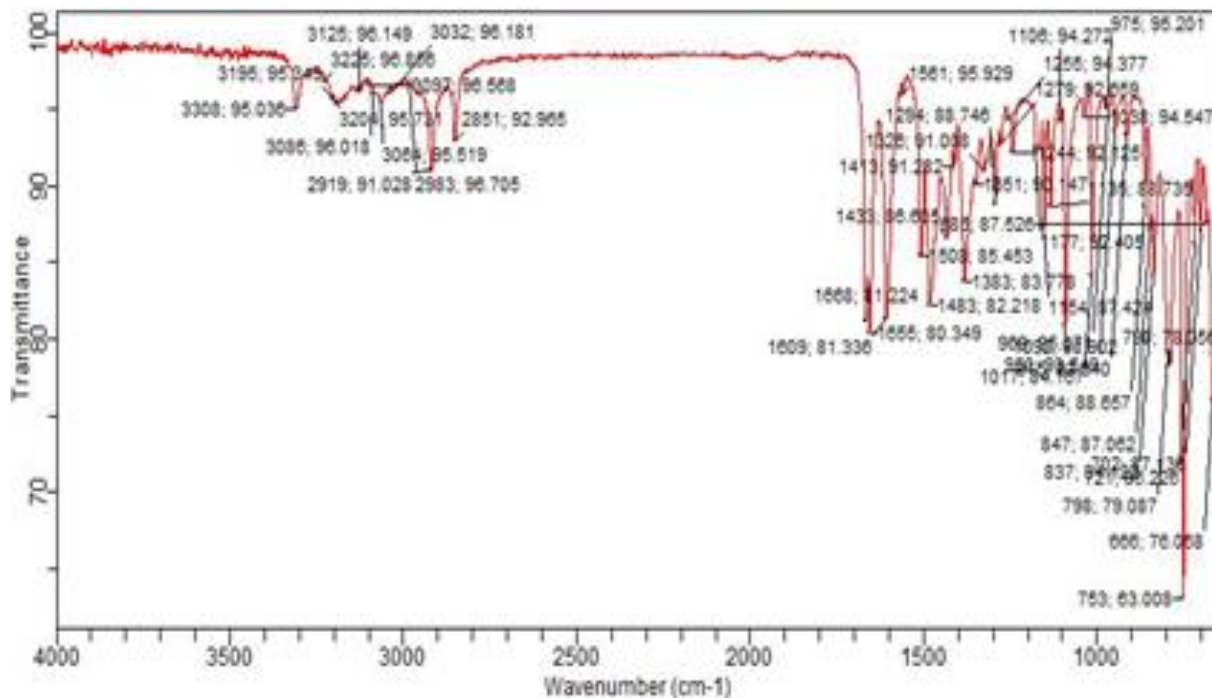


Figure 1 FTIR spectra of Flecainide

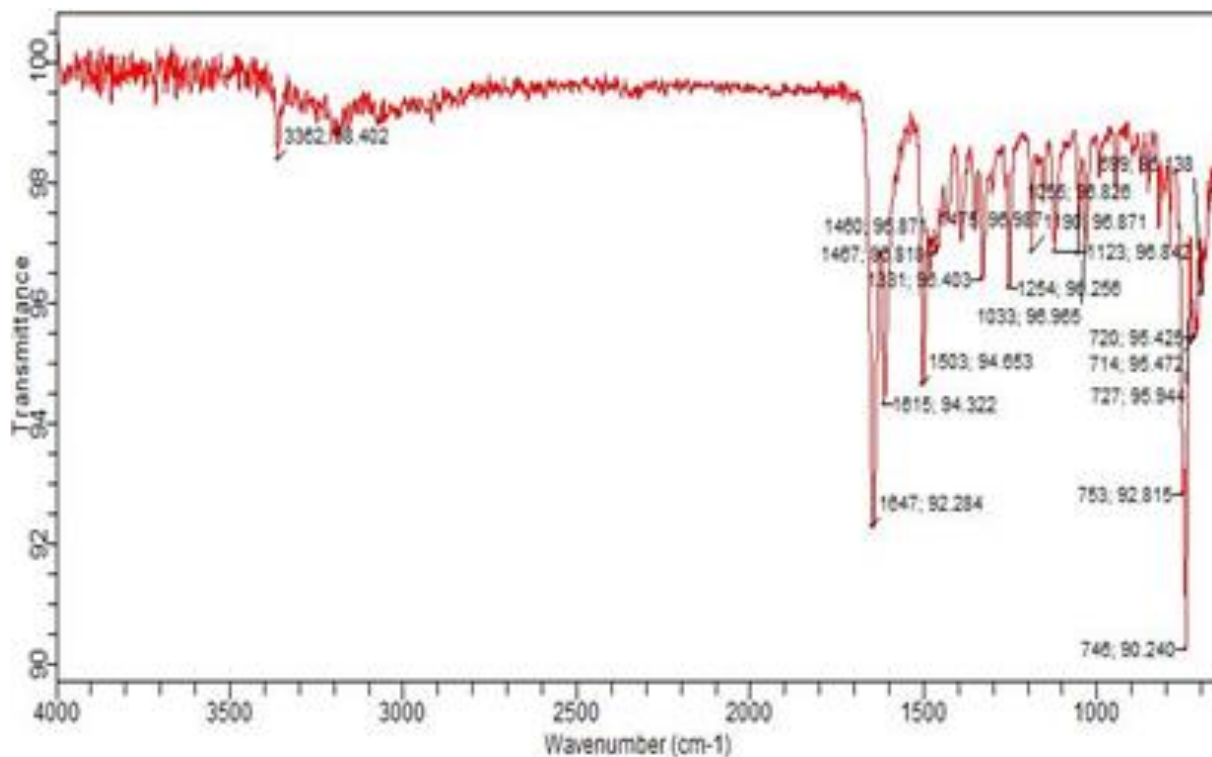


Figure 2 FTIR Spectrum of Mixture of compounds

peaks were present in the combination spectrum, demonstrating the drug and polymer's compatibility. IR spectra of each polymer and Cefotaxime combination with each polymer are displayed in **Figures 1** and **Figure 2**, using data from **Table 2**.

Table 2 FTIR interpretation data of Drug and Mixture of Compounds

Functional Groups	Flecainide	Mixture of compounds
(s)= C-H bend (Alkenes)	1000-650	1000-650
(m)O-H bend (Carboxylic Acid)	950-910	3300-2500
(s, b) N-H wag (1°,2° amines)	910-665	910-665
(m) C-Br stretch (Alkyl halides)	725-720	1300-1150
(s)C-O stretch (Alcohol, carboxylic acid, esters, ethers)	1320-1000	1320-1000
(m)C-N stretch (Aliphatic amines)	1250-1020	1250-1020
(m) C-C stretch (in-ring) (Aromatics)	1500-1400	3100-3000

Evaluation Parameters

Surface Morphology

SEM was used to examine the Flecainide beads' surface morphology. SEM images displaying the enhanced mixture. Surface smoothness was observed with guar gum incorporated Flecainide beads.

Frequency distribution analysis

The mean particle size of the Flecainide beads also decreased when the polymer ratio increased (**Table 3**). The rise in droplet viscosity could be the cause of the notable decrease. Obtaining flecainide beads with a normal frequency distribution and a size range of 1. to 1. mm was possible.

Percentage yield

The % practical yield of flecainide hydrogel beads has been calculated to determine the efficiency or percentage yield for any given process, which aids in selecting the most suitable manufacturing technique. The weight of the flecainide beads recovered from each batch divided by the total starting material was used to compute the practical yield, as shown in **Table 3**.

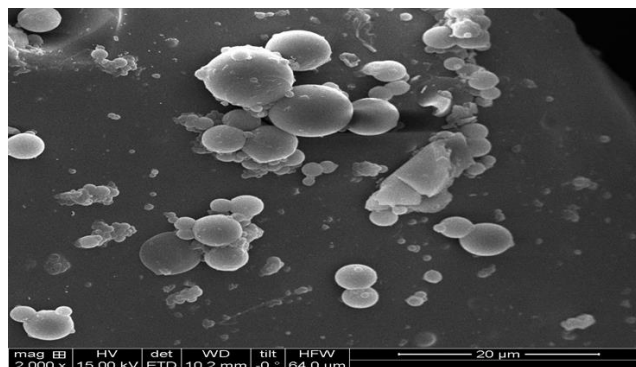


Figure 3 SEM photographs of hydrogel beads

Drug Content

As the polymer concentration increased, the drug content correspondingly increased. The data suggest that the distribution of Flecainide in the beads is appropriate, and the deviations are within allowable bounds, as Table 3 demonstrates.

Percentage of drug entrapment efficiency

As the concentration of polymer increased, its entrapment efficiency correspondingly increased. According to the findings, Flecainide has an appropriate distribution in the beads, and any deviations are within allowable bounds. Increasing the polymer concentration enhanced encapsulation efficiency [**Table 3**]. The effectiveness of trapping of highly concentrated beads made with carbopol.

In vitro dissolution studies

The Flecainide hydrogel beads' in vitro performance demonstrated a prolonged and regulated release of Flecainide. The in vitro dissolution trials' outcomes demonstrated predictable, regulated release. It was discovered that the medication release from the hydrogel beads decreased as the polymer content rose. Carbopol more successfully delayed drug release

Table 3 Average particle size of Flecainide Hydrogel beads

Formulation code	Average size (mm)	Percentage Yield	Entrapment efficiency (%)	Drug Content (%)
F1	1.1	83.32	73.23	75.42
F2	1.3	86.16	67.12	65.32
F3	1.4	88.41	81.78	79.38
F4	1.1	83.19	77.85	82.36
F5	1.2	85.17	82.83	89.43
F6	1.4	89.38	85.61	87.67
F7	1.2	88.49	77.54	90.32
F8	1.4	91.54	84.21	95.76
F9	1.1	92.66	83.24	94.34
F10	1.2	93.45	77.14	96.68
F11	1.2	94.32	89.18	97.77
F12	1.2	95.41	97.19	98.82

than HPMC K4M and sodium CMC; hydrogel beads reached their maximum release by the end of the 12th hour. **Table 4** and **Figure 4** exhibit every formulation's in vitro release profiles (F1 to F12).

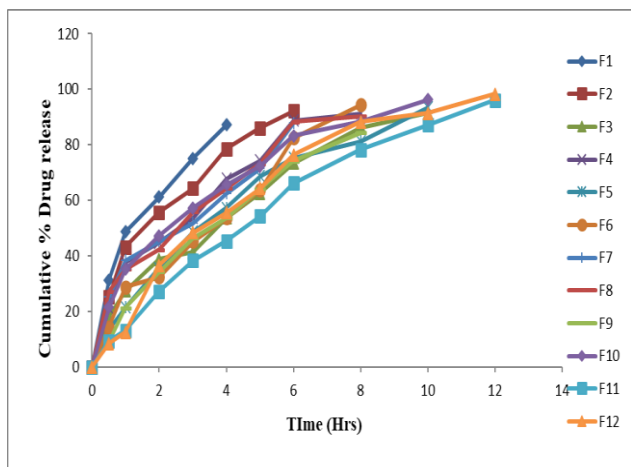


Figure 4 Flecainide hydrogel beads made of sodium alginate: in vitro release data

Release Order Kinetics of Flecainide Hydrogel Beads

The best formulation F9 in vitro dissolution data were fitted using various kinetic models, including the zero order, first order, Higuchi, and Korsmeyer-Peppas equations. It is confirmed as it follows the zero-order release because its value is closer to the '1' [Table 9]. The Korsmeyer and Peppas plot

[Figure 5, Figure 6, Figure 7, Figure 8] confirms the drug release mechanism.

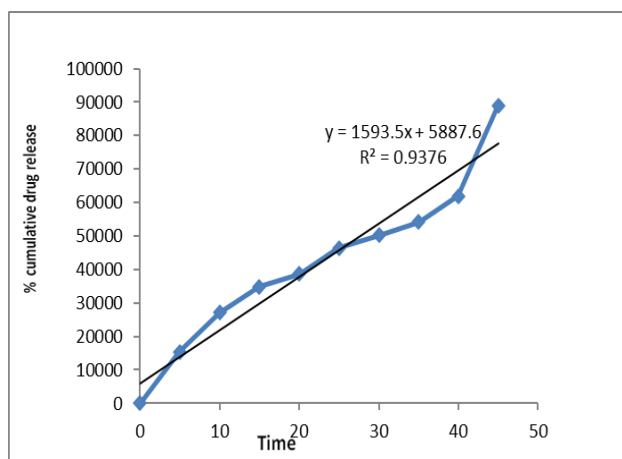


Figure 5 F12 of Zero Order Kinetics In vitro Dissolution Studies

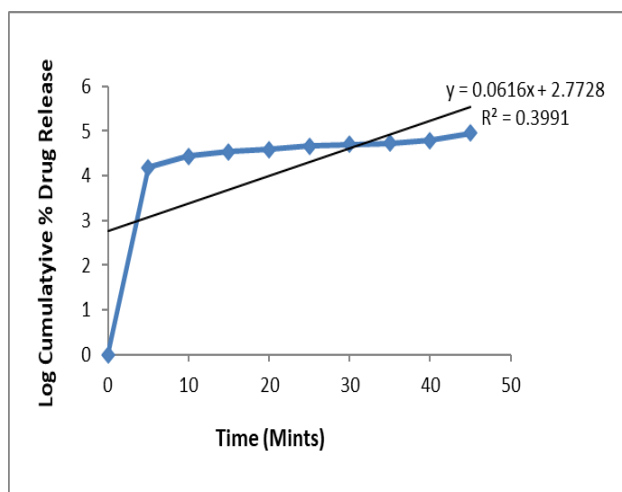


Figure 6 F12 of First Order Kinetics In vitro Dissolution Studies

Table 4 Flecainide hydrogel beads made of sodium alginate: in vitro release data

TIME (hrs)	F1	F2	F3	F4	F5	F6	F7	F8	F9	F10	F11	F12
0	0	0	0	0	0	0	0	0	0	0	0	0
0.5	31.43	25.28	18.36	21.32	13.51	14.45	25.35	27.43	9.18	21.62	9.52	8.54
1	48.65	43.17	27.41	37.45	21.52	28.81	38.32	35.32	21.32	35.18	13.18	12.48
2	61.21	55.81	38.91	44.82	35.81	32.51	45.32	42.32	34.43	47.32	27.34	36.49
3	75.15	64.31	41.42	53.71	48.72	45.33	51.85	55.75	46.51	57.28	38.44	48.52
4	87.31	78.64	53.87	67.88	57.34	53.65	62.51	64.34	53.56	65.58	45.38	55.61
5	-	86.22	62.61	74.34	68.81	63.88	71.65	73.34	64.32	72.31	54.32	64.42
6	-	92.25	73.56	88.87	75.33	82.34	87.92	88.32	74.32	83.33	66.34	76.42
8	-	-	86.21	91.41	81.33	94.54	-	90.44	84.43	88.32	78.26	88.28
10	-	-	91.32	-	93.53	-	-	-	-	96.32	87.29	91.22
12	-	-	-	-	-	-	-	-	-	-	96.18	98.31

Table 5 Drug Release Kinetics.

Batch	Zero Order	First Order	Higuchi	Peppas	Peppas
Code	r ²	r ²	r ²	r ²	n
F1	0.966	0.822	0.948	0.623	1.032
F2	0.978	0.816	0.949	0.632	1.022
F3	0.976	0.822	0.966	0.612	1.023
F4	0.962	0.843	0.964	0.633	1.032
F5	0.951	0.811	0.987	0.621	1.018
F6	0.962	0.843	0.945	0.624	1.013
F7	0.964	0.832	0.963	0.618	1.017
F8	0.977	0.811	0.977	0.617	1.018
F9	0.967	0.388	0.888	0.776	1.012
F10	0.979	0.812	0.945	0.637	1.021
F11	0.973	0.824	0.963	0.615	1.022
F12	0.841	0.841	0.961	0.974	1.021

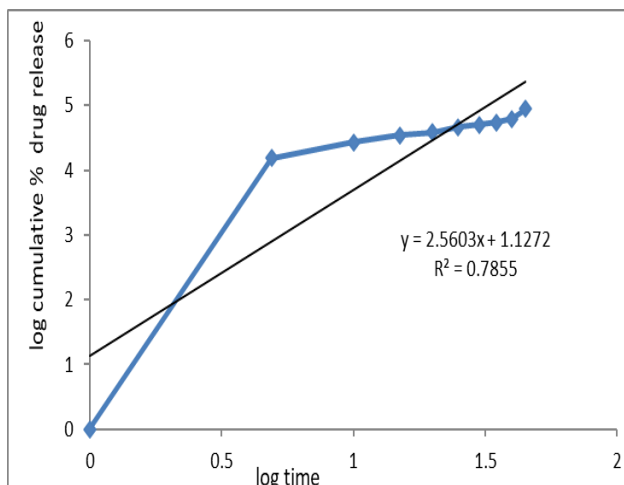


Figure 7 F12 of the Korsmeyer peppas in vitro dissolution studies

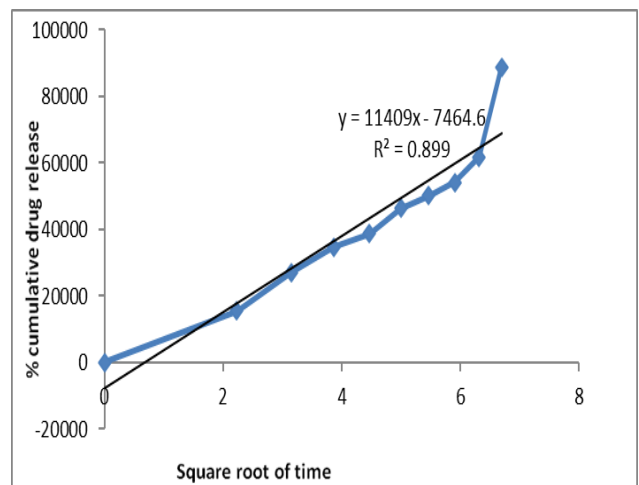


Figure 8 F12 of the Higuchi in vitro dissolution studies

STABILITY STUDY:

Formulation optimized for one to three months, F12 underwent stability trials, and the tablets' pharmacological content was examined. The outcomes are shown in **Table 6**.

Table 6 Studies on the stability of the optimized F12 formulation

Time in hrs	Drug Content		
	F12	After 1 Month	After 3 Month
1	75.43	75.41	74.31
2	65.43	65.31	64.31
3	79.22	79.14	80.38
4	82.24	82.43	81.61
5	89.41	89.45	90.43
6	87.28	87.14	86.46
7	90.27	90.32	91.65
8	95.67	94.34	94.61
9	94.34	93.25	93.54
10	96.49	95.88	95.68
11	97.32	96.77	96.32
12	98.67	97.77	96.79

CONCLUSION

Studies conducted before formulation, such as solubility and UV analysis, met the requirements. Flecainide and polymers did not interact, according to the FTIR Spectra. By using SEM, the flat surface of the Flecaïnide beads was verified. The Flecaïnide hydrogel beads' mean particle size decreased as the polymer ratio increased. Ordinary frequency distribution hydrogel beads containing fluorescein were produced. As the concentration of polymer developed, its entrapment efficiency correspondingly increased. The results suggest that the distribution of Flecaïnide in the beads was appropriate and that the variation remained within allowable bounds. The research findings also indicate that an increase in polymer concentration leads to a decrease in drug release. The Flecaïnide Hydrogel beads' in vitro performance demonstrated a sustained and regulated drug release. The in vitro dissolution data for the optimal formulation F12 were fitted using various kinetic models, including the Higuchi, Korsmeyer-Peppas, zero-order, and first-order equations. The optimized formulation F12 displays a 0.974 r^2 value. It conforms when approaching the zero-order release since its value

is closer to the '1'. The scenario involving Korsmeyer and Peppas further confirms the drug release mechanism. When the 'n' value for the optimized formulation (F12) is 1.021, the n value was more significant than 0.89, indicating Super case transport.

ACKNOWLEDGEMENT:

We want to express our profound appreciation to our management and principal, Dr. Y Prapurna Chandra from Ratnam Institute of Pharmacy, Pidathapolur (V & P), Muthukur (M), SPSR Nellore District, for their support, without which it would not have been possible to complete our research work.

Conflict of Interest

The authors declare no conflict of interest, financial or otherwise.

Funding Support

The authors declare that they have no funding for this study.

REFERENCES

- [1] Malpure P S, Patil S S, More Y M, Nikam P P, et al. A Review On-Hydrogel. American Journal of Pharmaceutical Research. 2018;8(3): 2249-3387.
- [2] Bindu SM, Ashok V, Chatterjee A, et al. As a Review on Hydrogels as Drug Delivery in the Pharmaceutical Field. International Journal of Pharmaceutical and Chemical Science. 2012;1(2).
- [3] Ullah F, Hafi Othman M B, Javed F, Ahmad Z, Akil H M, et al. Classification, Processing, and Application of Hydrogels: A Review, Materials Science & Engineering C. 2015;57:414-433.
- [4] N Das et al. Preparation Methods and Properties of Hydrogel: A Review. International Journal of Pharmaceutical Sciences. 2013;5(3):112-117
- [5] Kim B, Peppas N A, et al. Poly (ethylene glycol)-Containing Hydrogel for Oral Protein Delivery

- Applications, Biomed. Microdevices. 2003;5(4):333-341.
- [6] Richter A, Paschew G, Klatt S, Lienig J, Arndt K F, Adler H J P, et al. Review on Hydrogel-based pH Sensors and microsensors. *Sensors*. 2008;8(1):561-581.
- [7] Na K, Lee K H, Bae Y H, et al. pH-sensitivity and pH-dependent Interior Structural Change of Self-assembled Hydrogel Nanoparticles of Pullulan Acatate/Oligo-sulfonamide Conjugate. *Journal of Controlled Release*. 2004;97(3):513-525.
- [8] Tabata Y, Miyao M, Ozeki M, Ikada Y, et al. Controlled Release of Vascular Endothelial Growth Factor by Use of Collagen Hydrogels. *J. Biomater. Sci. Polymer Edn*. 2000;11(9):915-930.
- [9] Koetting M C, Peters J T, Steichen S D, Peppas N A, et al. Stimulus-responsive Hydrogels: Theory, Modern Advances, and Applications. *Materials Science and Engineering. R, Reports*. 2015;93:1-49.
- [10] Meshram PS, Kale SD, Labale PS, Mate KS, et al. Hydrogel Polymer: A Unique Material for Bio-Separation, Bio-Sensing, and Drug Delivery. *International Advanced Research Journal in Science, Engineering and Technology*. 2017;4(3):177-182.
- [11] Sing A, Sharma PK, Garg VK, and Garg G, et al. Hydrogels: a review. *International Journal of Pharmaceutical Sciences Review and Research*. 2010;4(2):97-105.
- [12] Sing SK, Dhyani A, and Juyal D, et al. Hydrogels: Preparation, characterization and Applications. *The Pharma Innovation Journal*. 2017;6(6):25-32.
- [13] Devi A, Nautiyal U, Kaur S, and Komal K, et al. Hydrogel: a smart drug delivery device. *Asian Pacific Journal of Health Sciences*. 2014;1(4S):92-105.
- [14] Vidya S, Sejal V, et al. Formulation and Evaluation of Microemulsion-Based Hydrogel For Topical Delivery. *International Journal of Pharmaceutical Investigation*. 2012;2(3):140-149.
- [15] Singh V, Singh PK, Sharm PK, et al. Formulation and Evaluation of Topical Gel of Aceclofenac Containing Piparine. *Indo-American Journal of Pharmaceutical Research*. 2013;3(7):5266-5280.

Copyright: This is an open access article distributed under the terms of the Creative Commons Attribution-Noncommercial- Share Alike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.



© 2024 Pharma Springs Publication